

Regulation of the Glycolytic Pathway

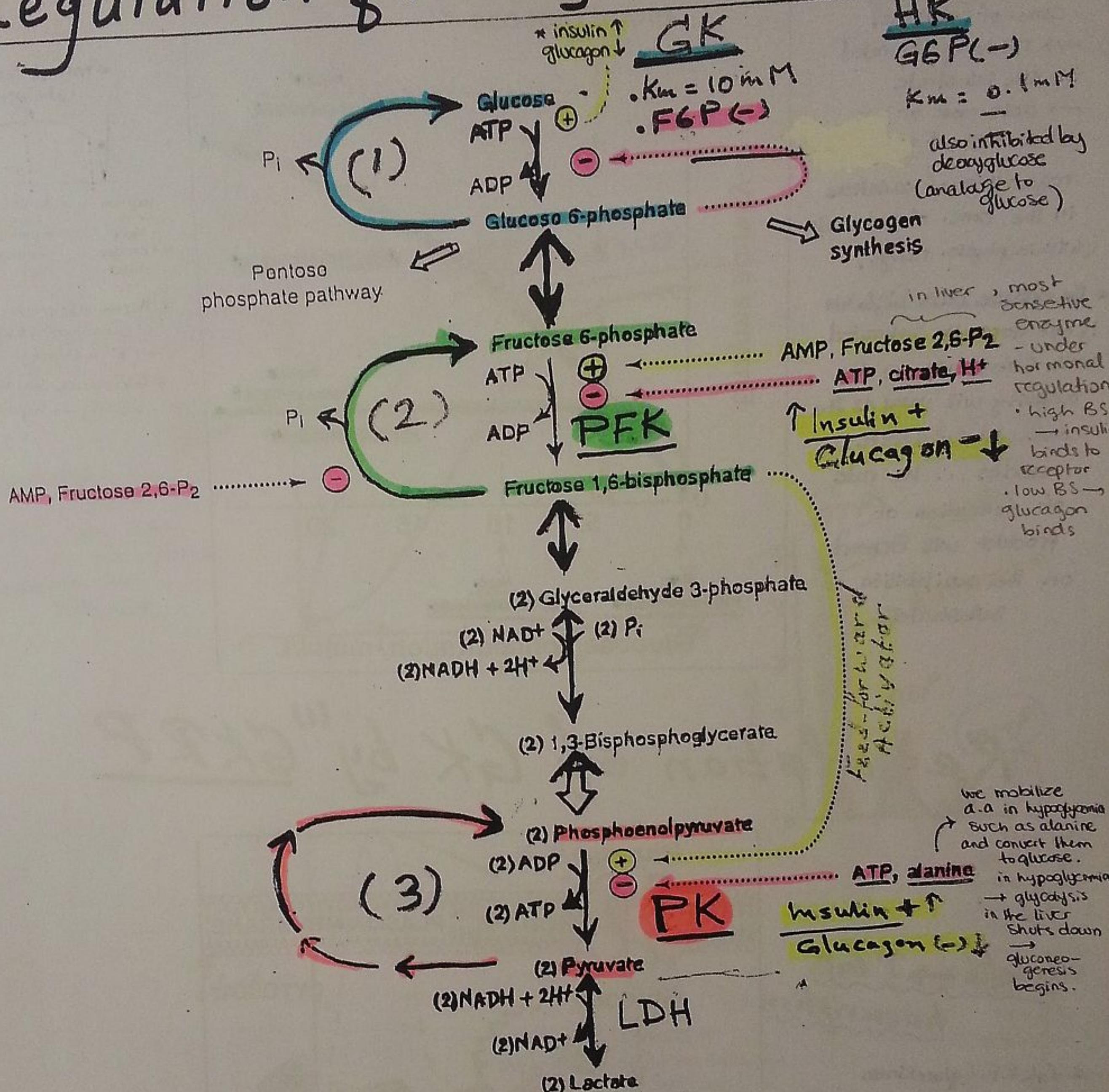


FIGURE 7.13

Important regulatory features of the glycolytic pathway.
Because of differences in isoenzyme distribution, not all tissues of the body have all of the regulatory mechanisms shown here.

* just to understand, own notes.

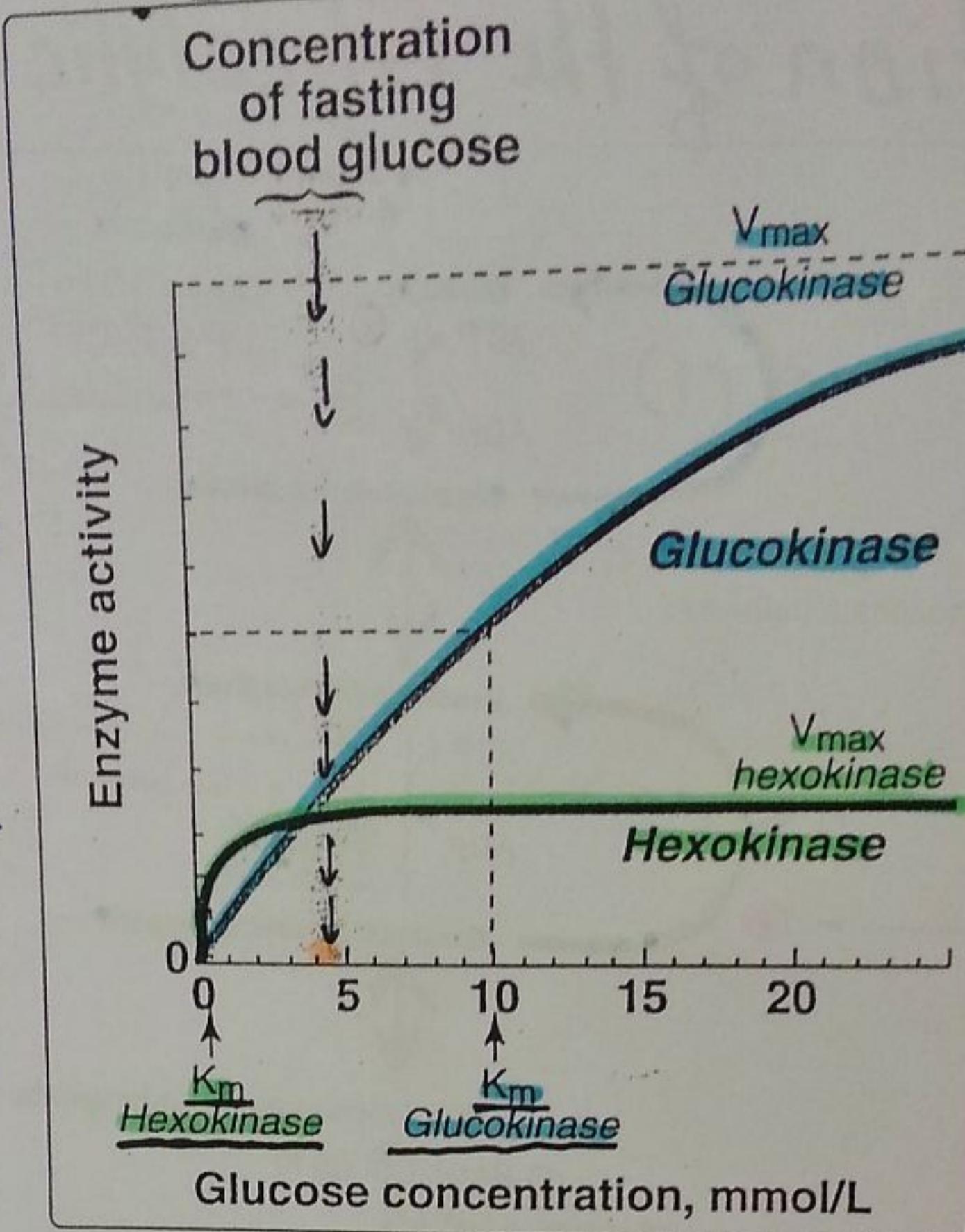
* note, *increased insulin \rightarrow increased GLUT-4 transporters \rightarrow rate of uptake of glucose for glycolysis increases, it upregulates the transcription of GK, PFK and PK levels.

* glucagon has the opposite effect, its secretion is stimulated by low blood glucose levels, promotes breaking down of energy storage molecules when energy is needed. down regulates the transcription of those three hormones

- V_{max} : rate of rxn when the enzyme is saturated with substrate = max rate of rxn
 - the relationship between the rate of rxn and conc. of substrate depends on the affinity of the enzyme to its substrate, expressed as K_m , an inverse measure of affinity.
- * high K_m \rightarrow low affinity \rightarrow requires a greater conc. of substrate to reach V_{max} .

Effect of Glucose

- * An enzyme with low K_m (relative to physio. conc. of substrate)
 - normally saturated with substrate
 - acts more or less at a constant rate, regardless of variations in the conc. of substrate (within physio. range)
- * An enzyme with high K_m is not normally saturated with substrate → its activity will vary as the conc. of substrate varies ∵ the rate of formation of product will depend on the availability of substrate.



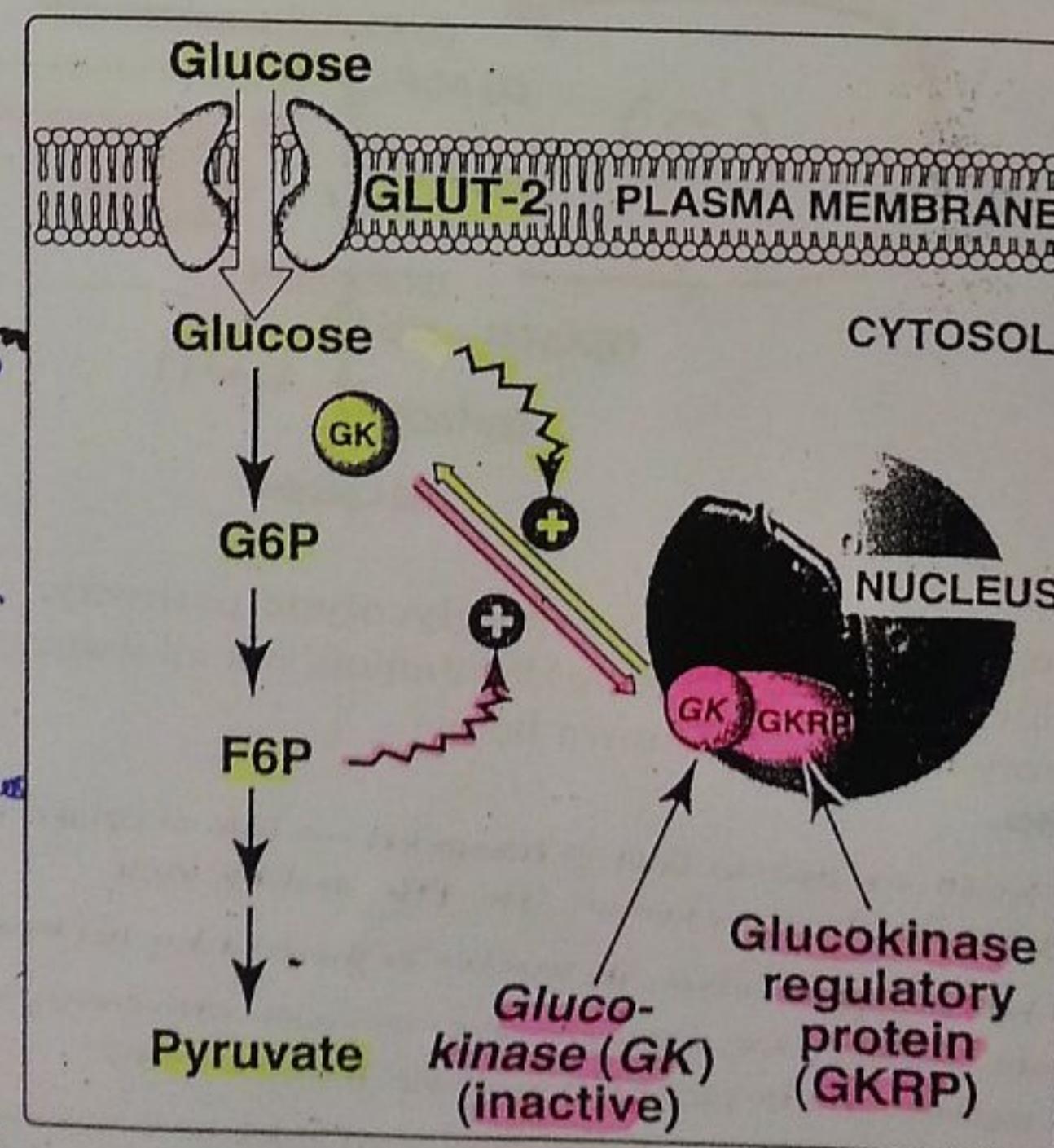
- * normal enzyme catalyzed on:
 - low sub. conc. → steep
 - high sub. conc. → plateau
 - enzyme unsat. → enzyme sat.
- * Hexokinase reaches saturation quickly → high affinity
- * Glucokinase has low affinity → higher V_{max}

Regulation of GK by "GKRP"

2)

insulin \rightarrow \uparrow GK transcription

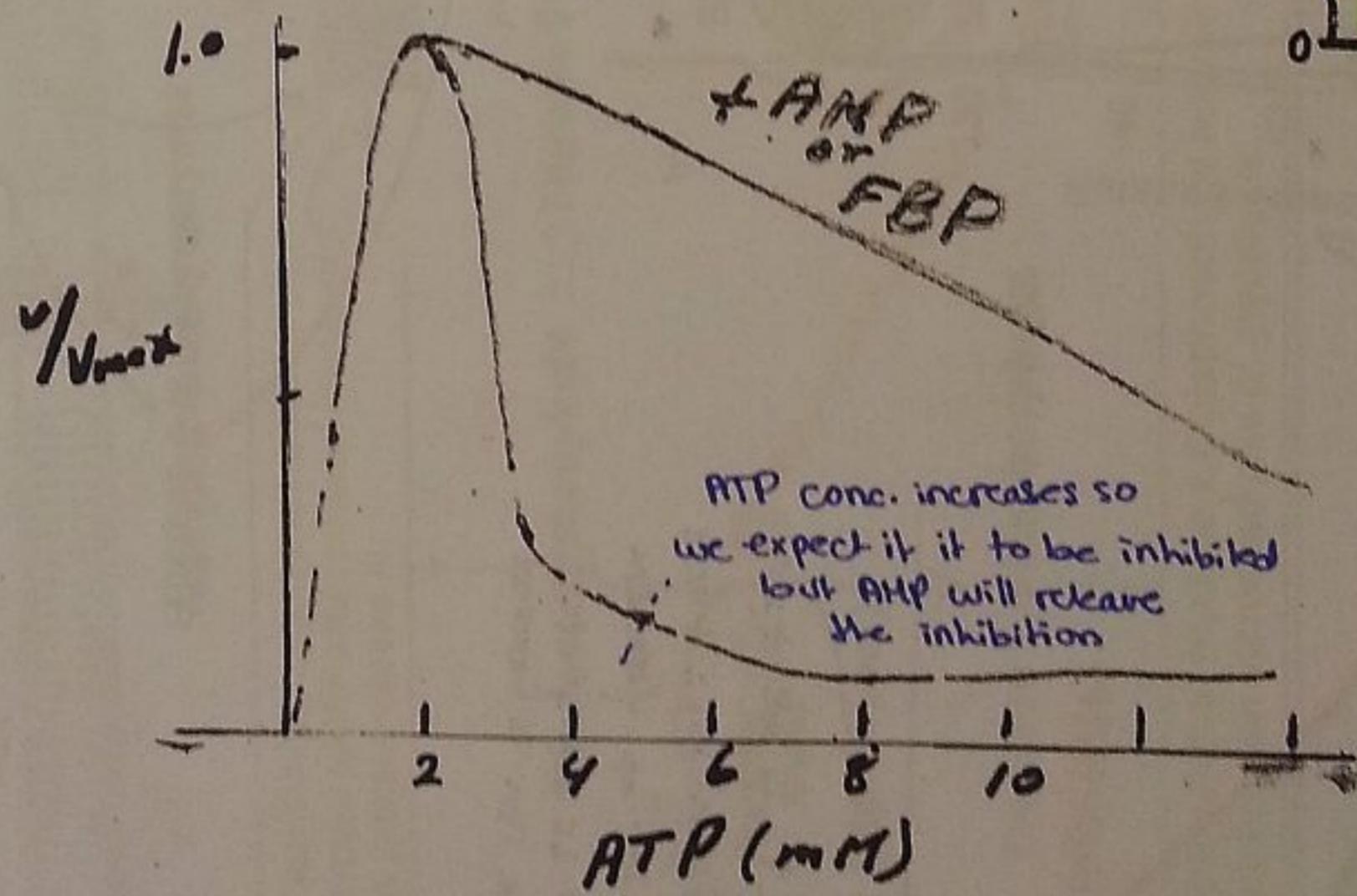
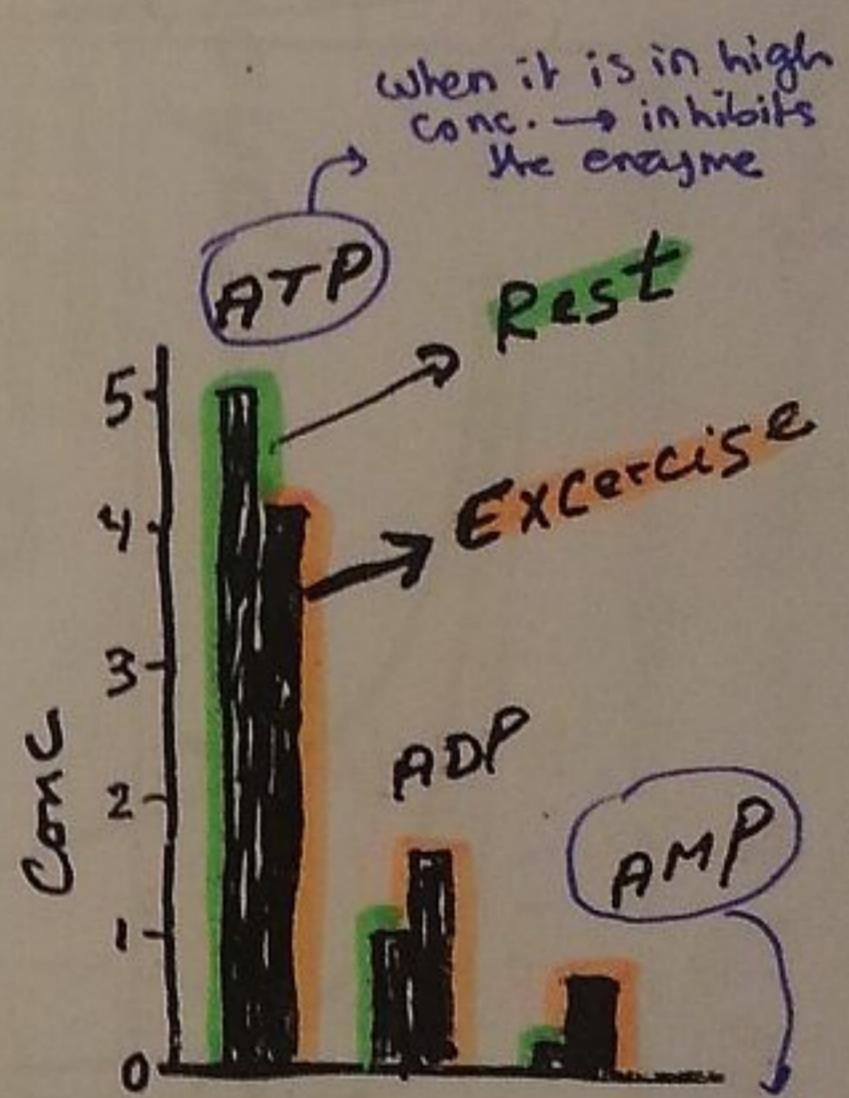
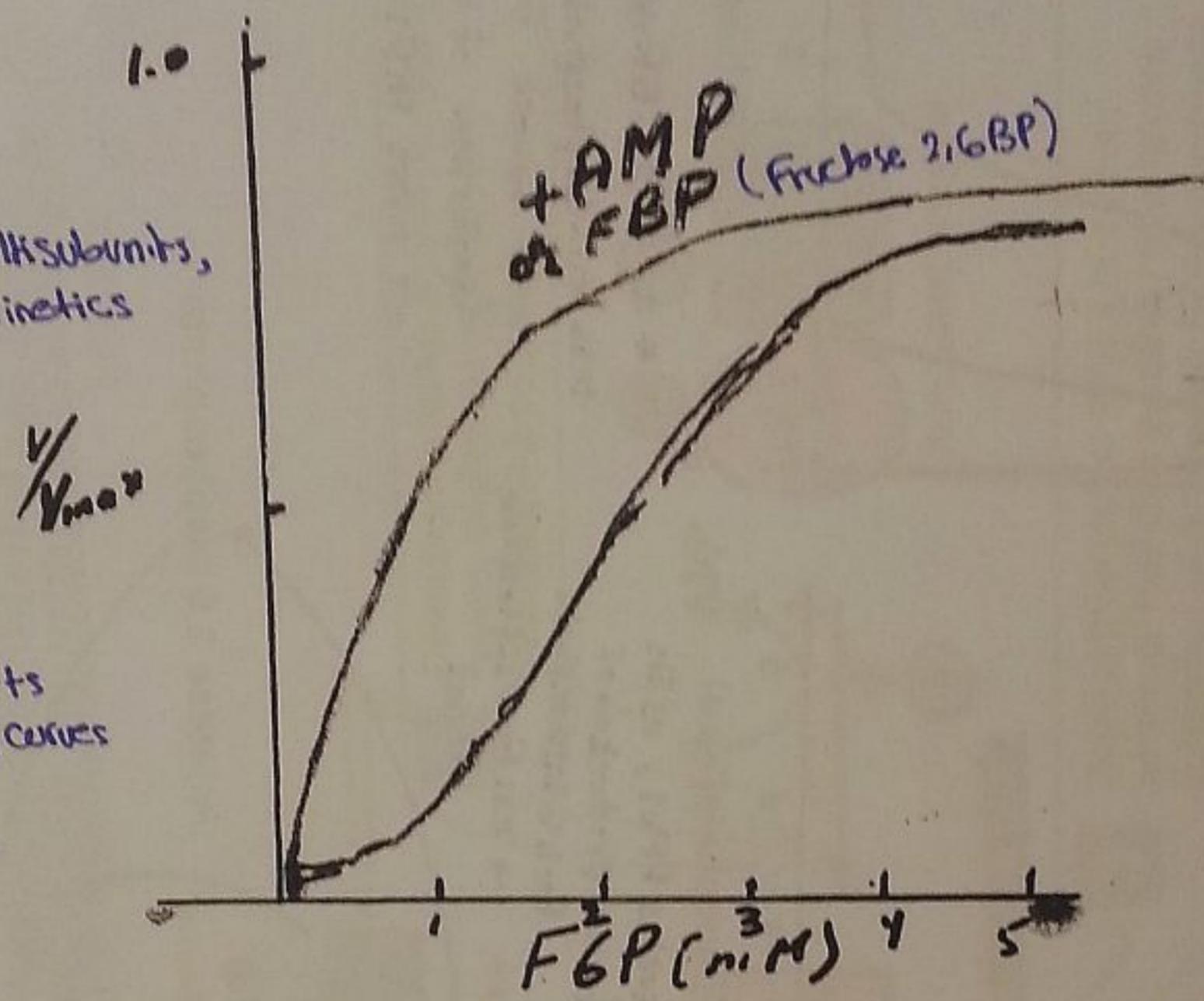
- * GKRP (glucokinase regulatory protein) in the liver regulates activity of GK through reversible binding.
 - Fructose 6-phosphate available
 - GK \rightarrow translocated into nucleus
 - GK binds tightly to GKRP
 - GK therefore is inactivated
 - When glucose levels in the blood (also in hepatocytes, due to GLUT-2) \uparrow
 - GK is released from GKRP
 - enzyme enters cytosol
 - Glucose is phosphorylated \rightarrow G6P.



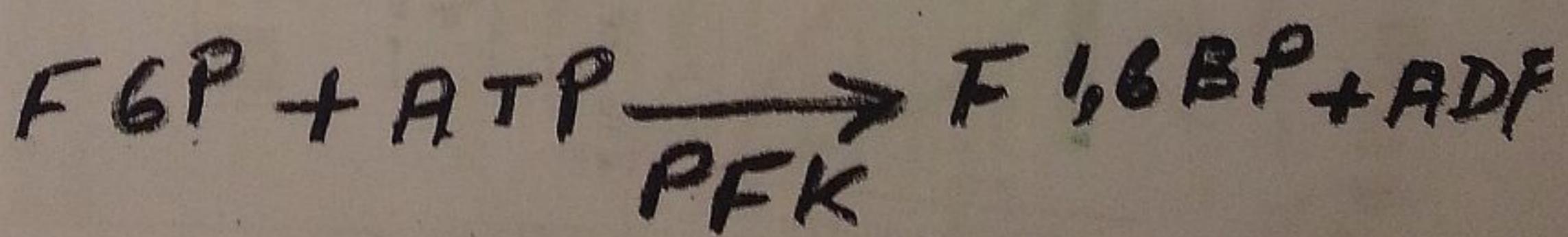
Regulation of PFK by AMP and ATP

* PFK consist of multisubunits, it shows sigmoidal kinetics with F6P.

* Sigmoidality can be abolished and the curve shifts to the left by various activators like AMP, FBP.



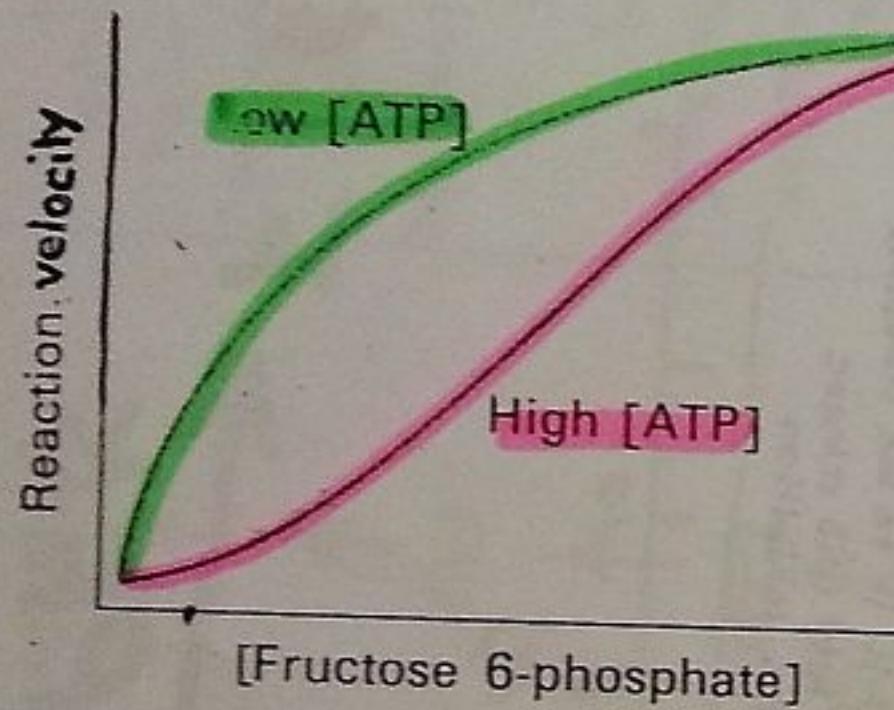
This inc. is enough to overcome the ATP inhibition → enzyme is activated during exercise (a de-inhibitor) releases inhibition by ATP.



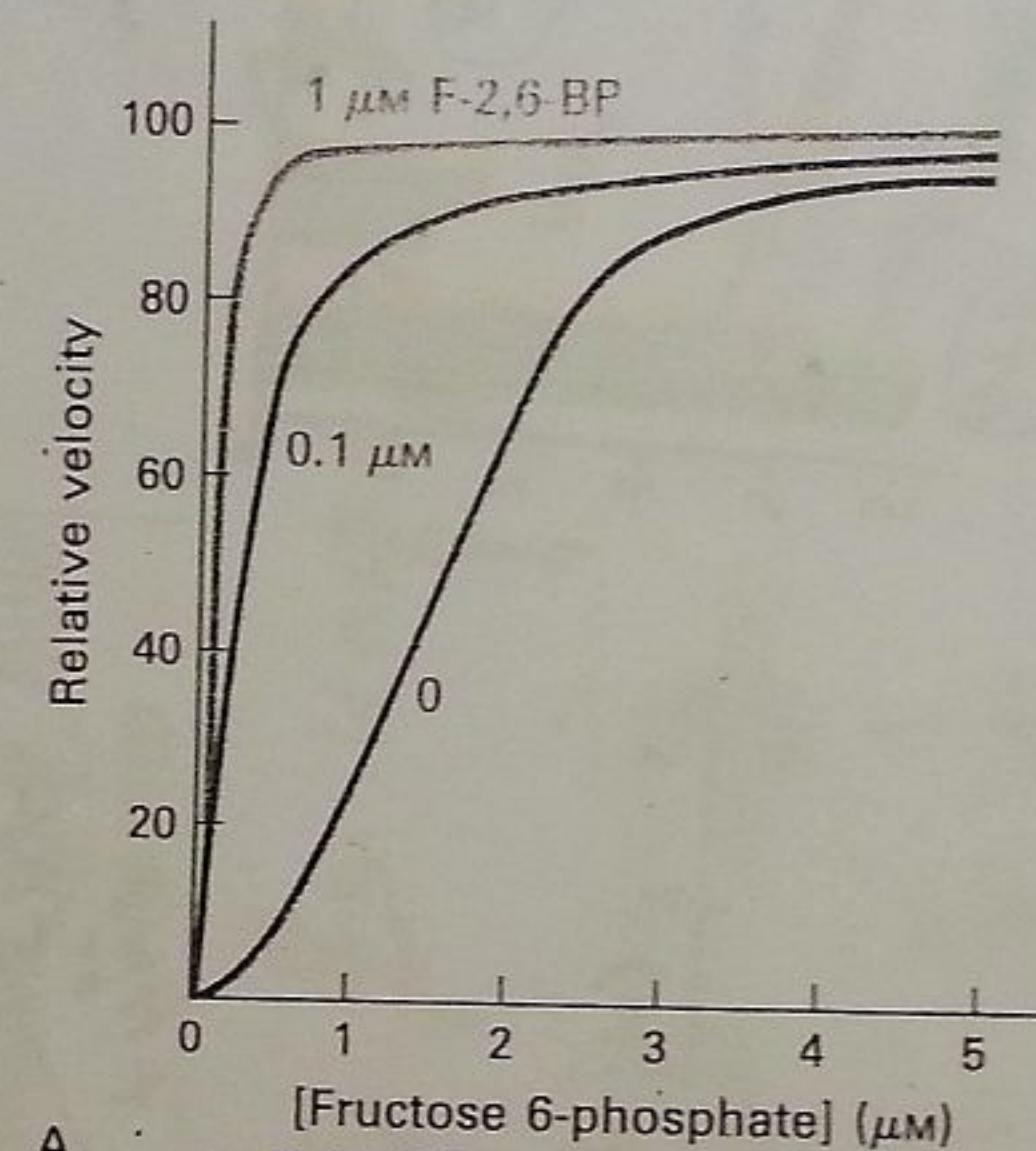
Regulation of PFK

* fru 2,6 BP:
* There is hormonal regulation

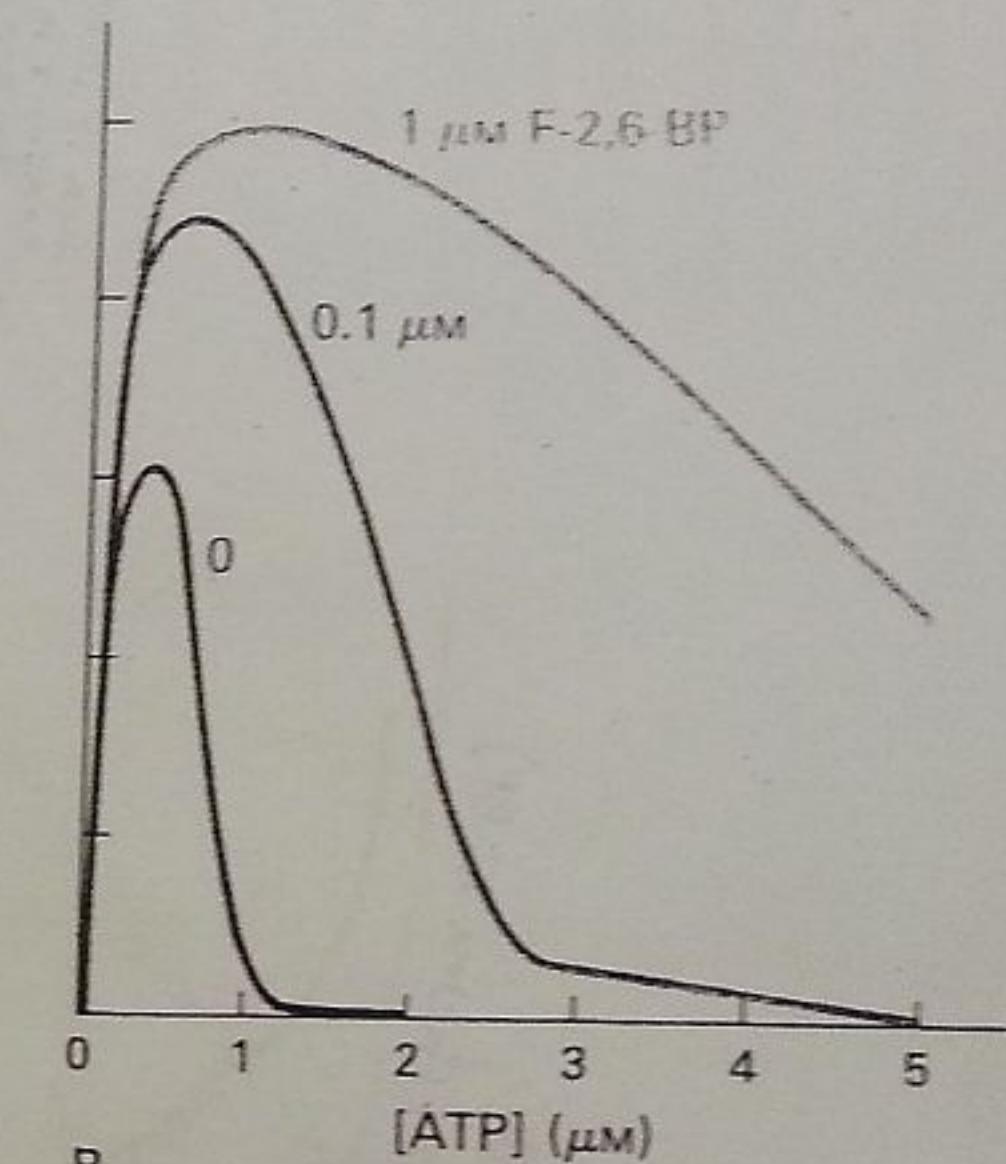
9a



* As ATP conc. ↑ → indicates a high rate of glycolysis ∵ it acts as an inhibitor → formation of free 1,6 bisphosphate ↓ → reverse rxn is favored



* F 2,6 BP activates Pfk1, as its conc ↑ → favors production of fru 1,6 bisphosphate
Stimulated by fructose 6-phosphate → less fru-6-phosphate formed



* as F-2,6-BP conc. ↑ → less fru-6-phosphate is formed since it favors forward rxn (production of fru 1,6 bisphosphate) → more ATP is produced in the process

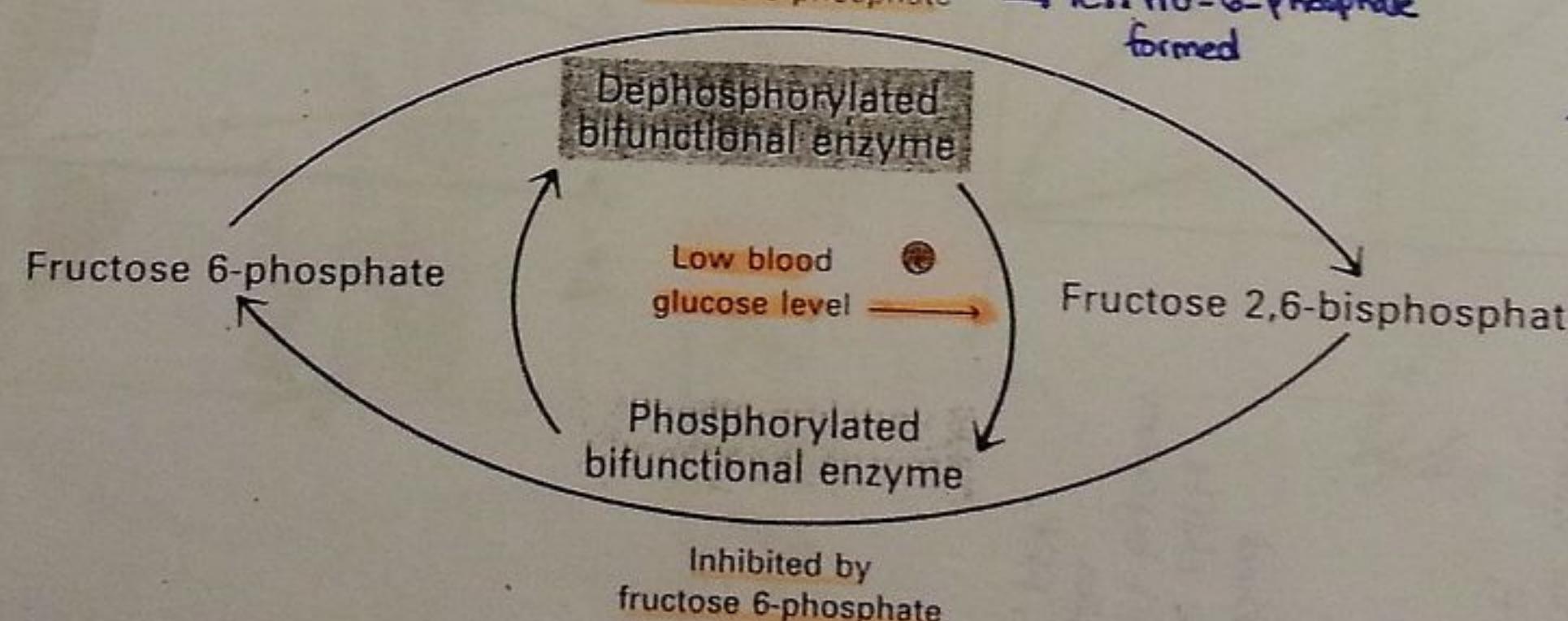
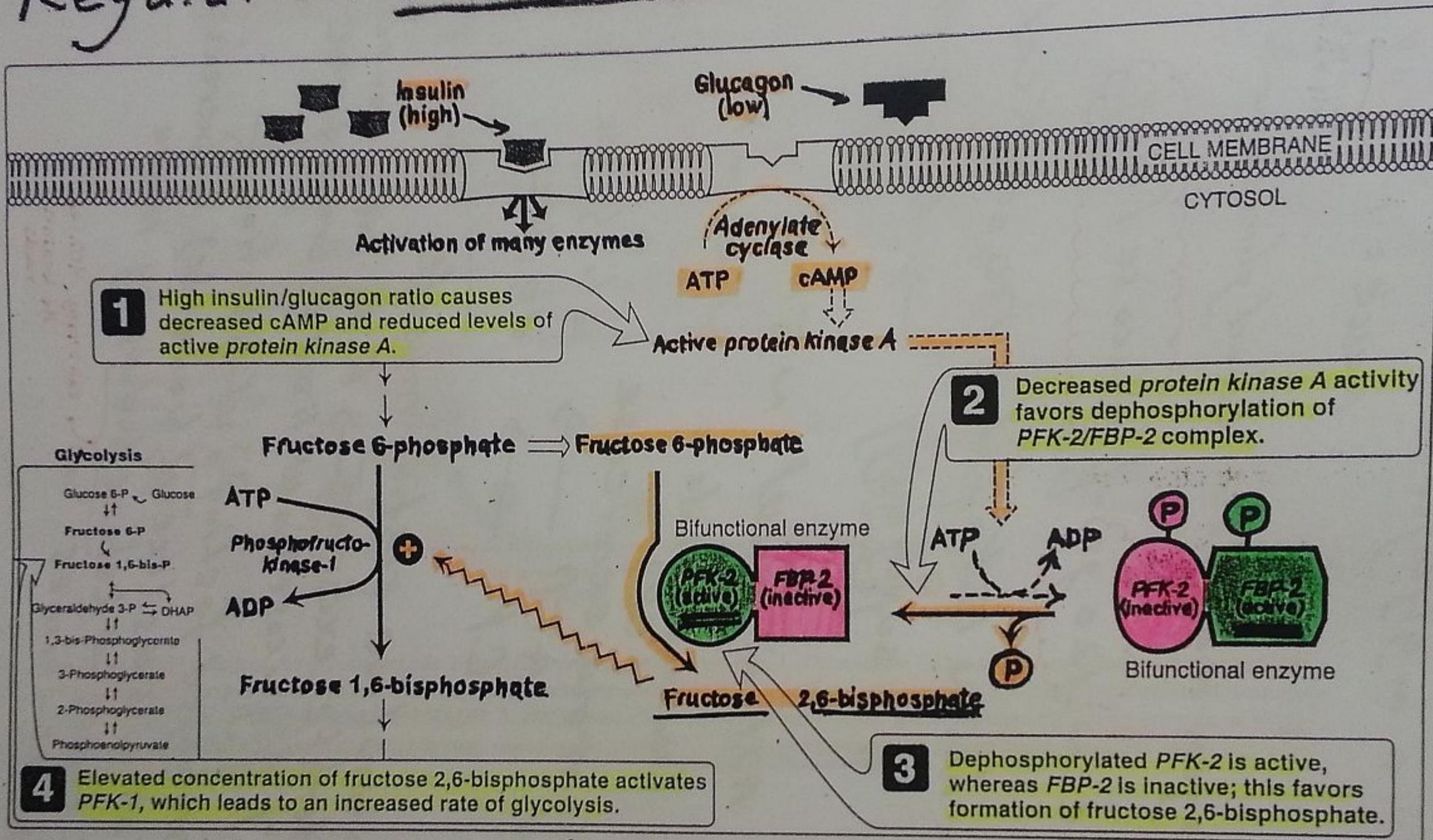


Figure 19-6, page 493; Figure 19-7, page 494; Figure 19-8, page 495

Regulation of Fru-2,6-BP Level



Regulation of PK by phosphorylation - Dephospha-

- * High insulin \rightarrow low cAMP \rightarrow PKA \downarrow \rightarrow Dephosphorylation of PFK-2 / FBP-2 complex \rightarrow dephosphorylated PFK-2 is active , FBP is inactive
 \Rightarrow formation of fructose 2,6 bisphosphate from fructose 6 phosphate
 \rightarrow fru 2,6 BP \rightarrow Activates PFK-1 \rightarrow glycolysis rate \uparrow

Distribution of P_Glucose:

Distribution of ~~150~~
free glucose in E.C.F. = 20 gr \equiv 80 k_{cal}

= anglycogen in liver = ~ 75 gr
= anglycogen in muscle = ~ 400 gr

glycogen in liver -
= muscle = ~ 400 gr
2% of net weight in liver

- liver glycogen maintains blood glucose \rightarrow 16 h

- Brain use ~ 120 gr of glucose / day

- Brain use $\sim 10\%$ of body energy
- 70 kg man has $\sim 15 \text{ Kg fat} \equiv 130,000 \text{ Kcal}$
 - " supply energy $\rightarrow 60 - 90 \text{ days}$

- conc of ATP in muscle $\sim 5 \text{ mM}$

" " Creatine phosphate = ~ 20 mM

" " (CP)
Upon vigorous exercise
ATP → 2 to 4 sec.
CP → 20 sec

- Post-absorptive resting muscle or with moderate exercise \rightarrow F. A. main source 80% of glucose is utilized by brain & rbc

- During prolonged fasting, utilization of F.A. by all tissues (except brain + rbc) is increased 4 to 5 times. Ketone bodies by more than 100-times
→ can supply 40% of the brain's needs

Glyceraldehyde 3-P dehydrogenase

converts G3P ^⑥ → 1,3BPG
aldehyde carb with carbon

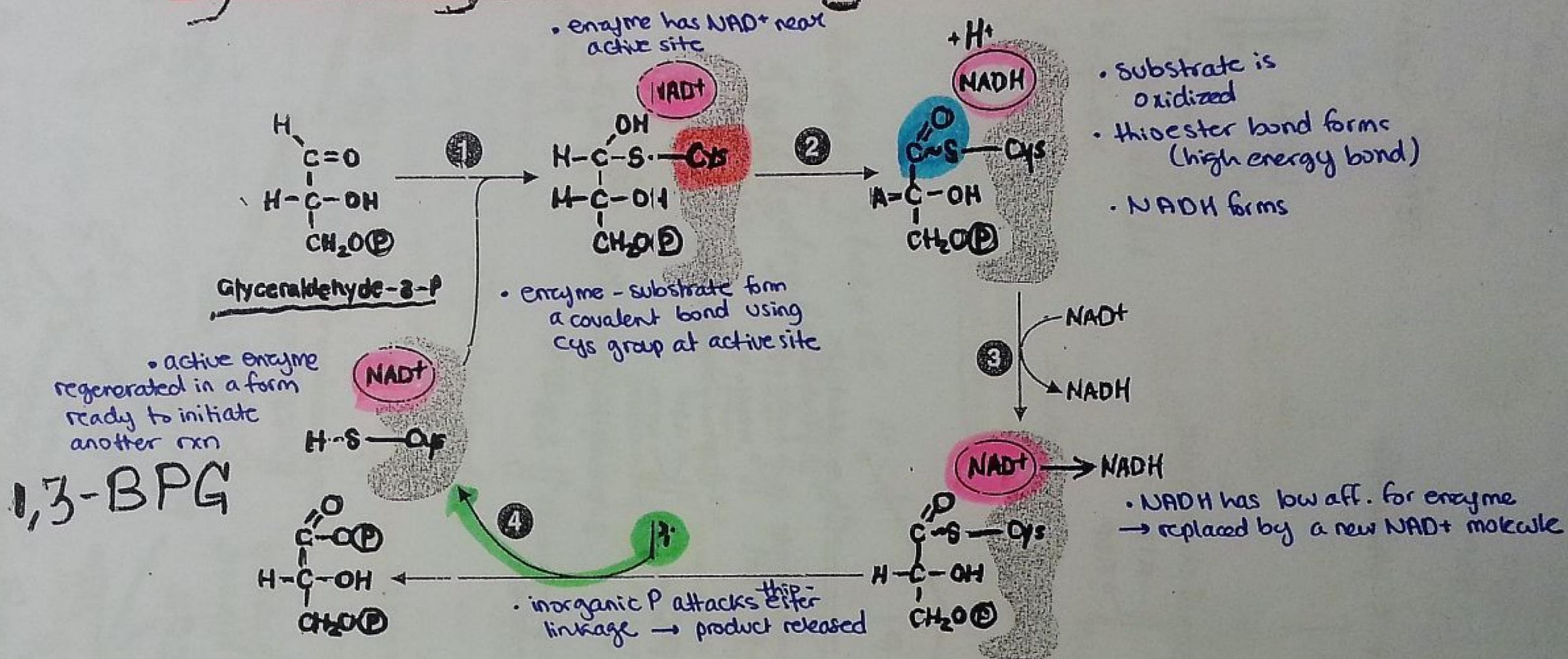


Fig. 22.17. Mechanism of the glyceraldehyde 3-phosphate dehydrogenase reaction. 1. The enzyme forms a covalent linkage with the substrate, using a cysteine group at the active site. The enzyme also contains bound NAD⁺ close to the active site. 2. The substrate is oxidized, forming a high-energy thioester linkage (in blue), and NADH. 3. NADH has a low affinity for the enzyme and is replaced by a new molecule of NAD⁺. 4. Inorganic phosphate attacks the thioester linkage, releasing the product 1,3 bisphosphoglycerate, and regenerating the active enzyme in a form ready to initiate another reaction.

Glycolysis Can be Inhibited

36 4

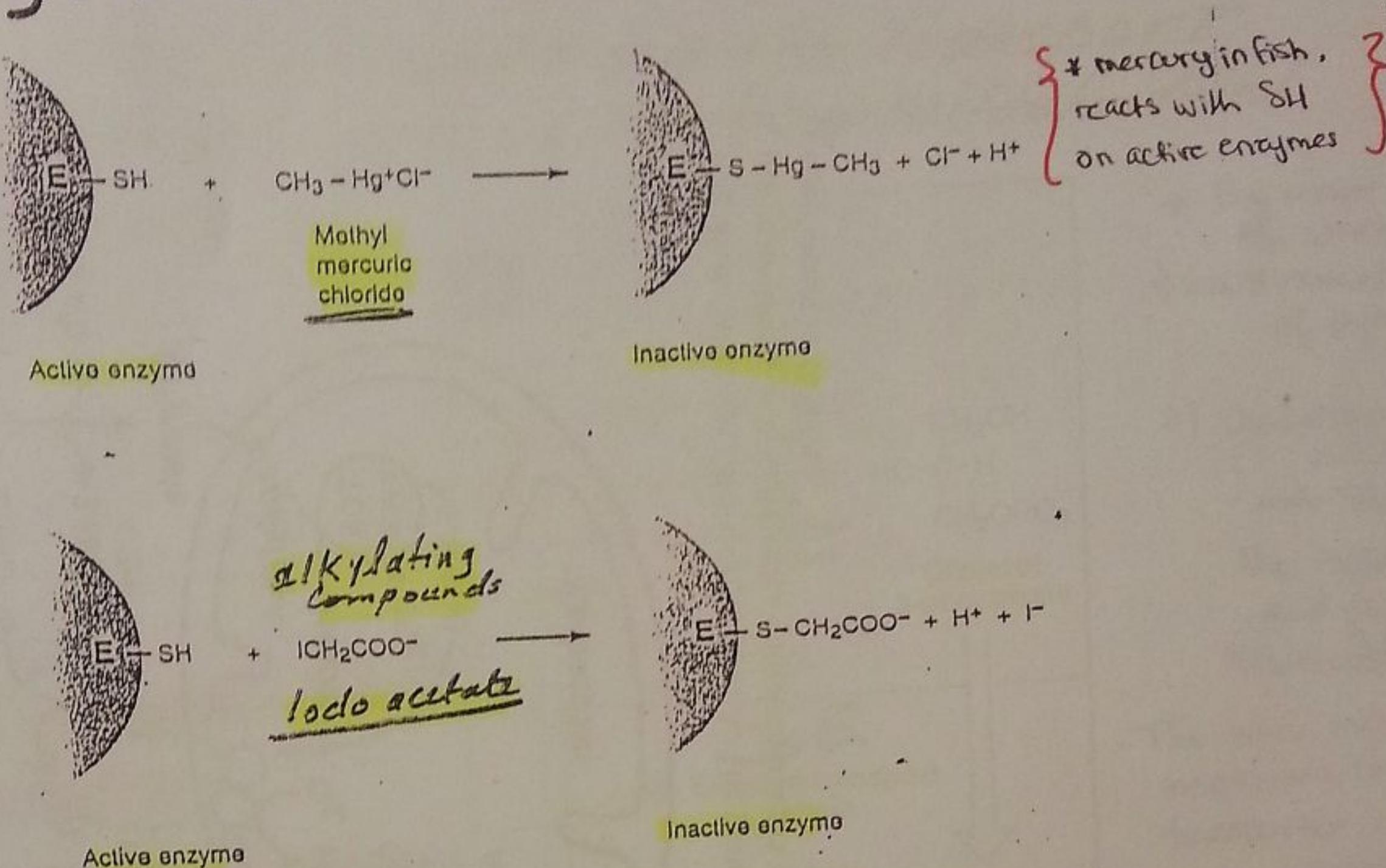
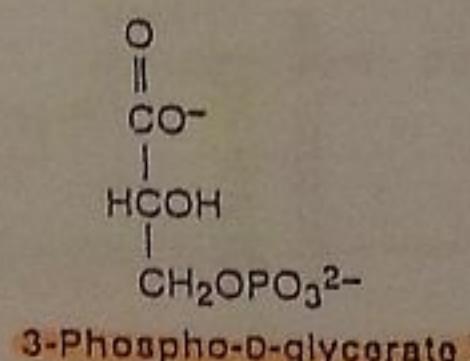
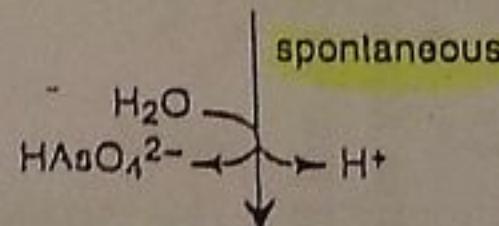
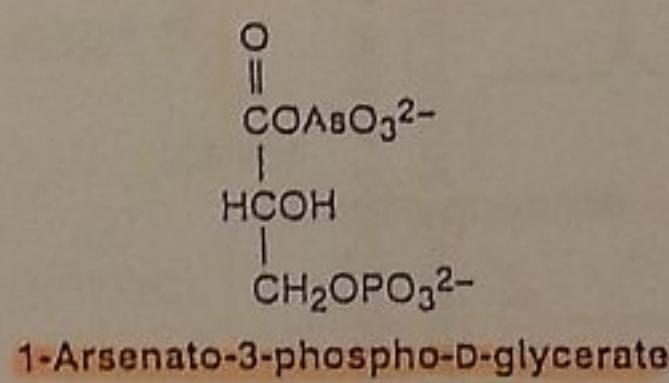
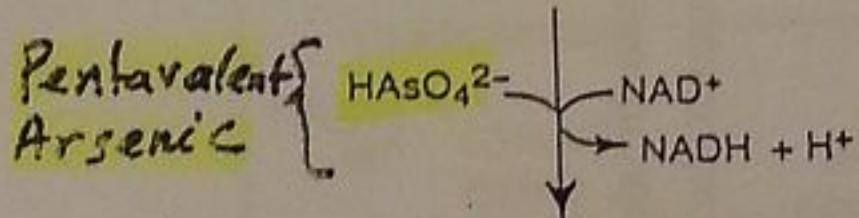
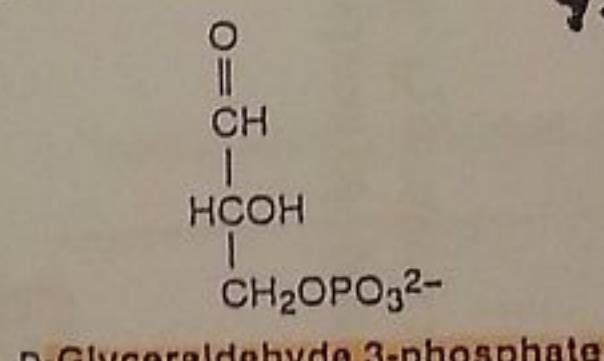


FIGURE 7.11
Mechanism responsible for inactivation of glyceraldehyde-3-phosphate dehydrogenase by sulfhydryl reagents.

- 1. 2-deoxy glucose $\text{HK} \downarrow$
- 2. Sulfhydryl Reagents $\text{GADH} \downarrow$
- 3. Fluoride Enolase \downarrow

* Pentavalent Arsenic (Arsenate) can prevent net ATP and NADH production:

- competes with Pi as a substrate for G3P
- 1-Arsenato-3-phosphoglycerate
- Hydrolyzes readily to form 3 Phosphoglycerate, bypassing the synthesis of 1,3 BPG → no ATP formed → cell is deprived of energy



4. Arsenate Inhibition HAsO_4^{2-}

Arsenolysis also interferes with ATP formation by Dk. Pi_i :

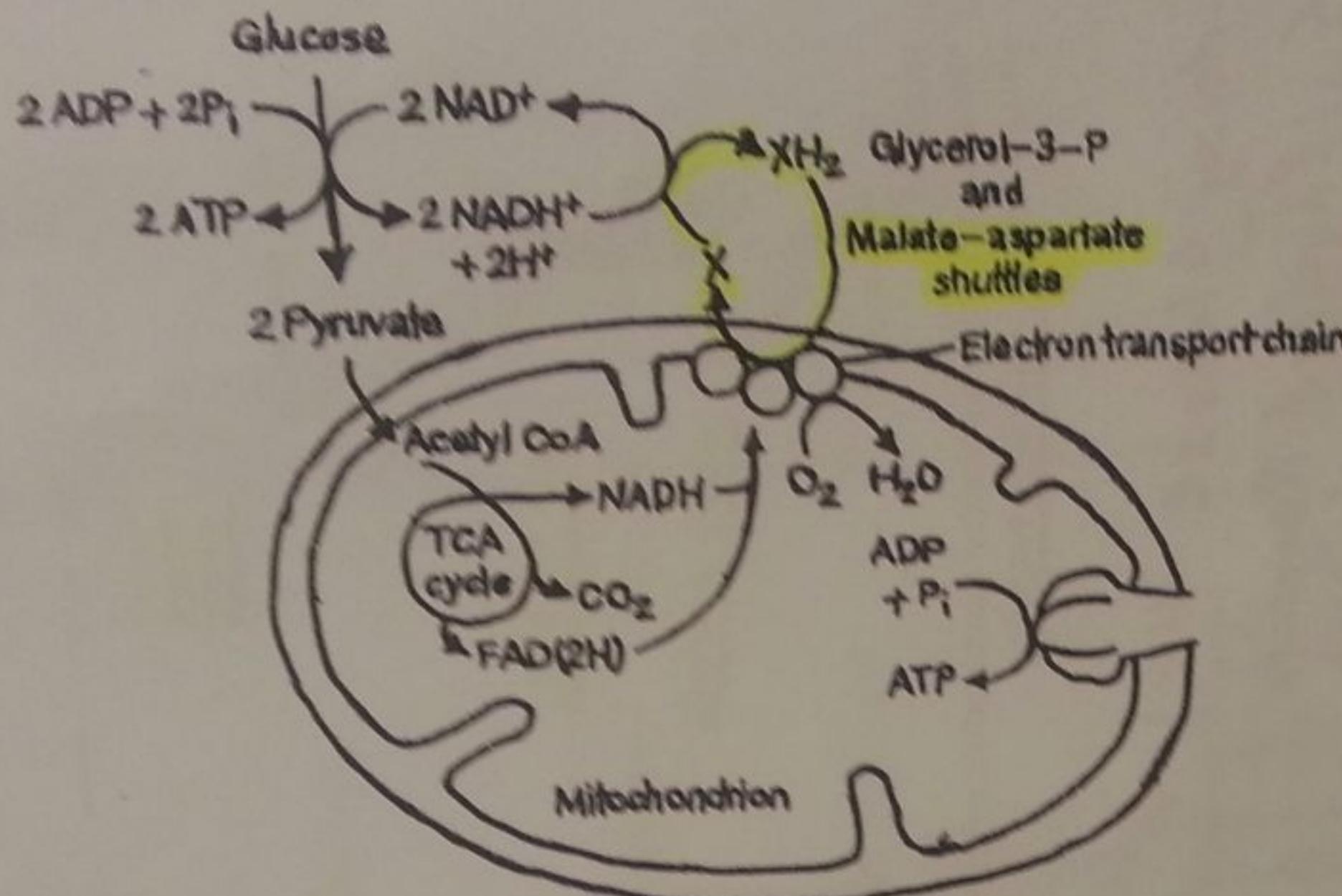
- can also replace phosphate in ATP
- replaces PO_4 (2 ATPs lost)

Arsenite (trivalent) AsO_3^{3-}

bind both -SH & the Cofactor lipoic acid and inhibiting e.g. PD, KG-D etc. more TOXIC

FIGURE 7.12
Arsenate uncouples oxidation from phosphorylation at the step catalyzed by glyceraldehyde-3-phosphate dehydrogenase.

A. Aerobic glycolysis



B. Anaerobic glycolysis

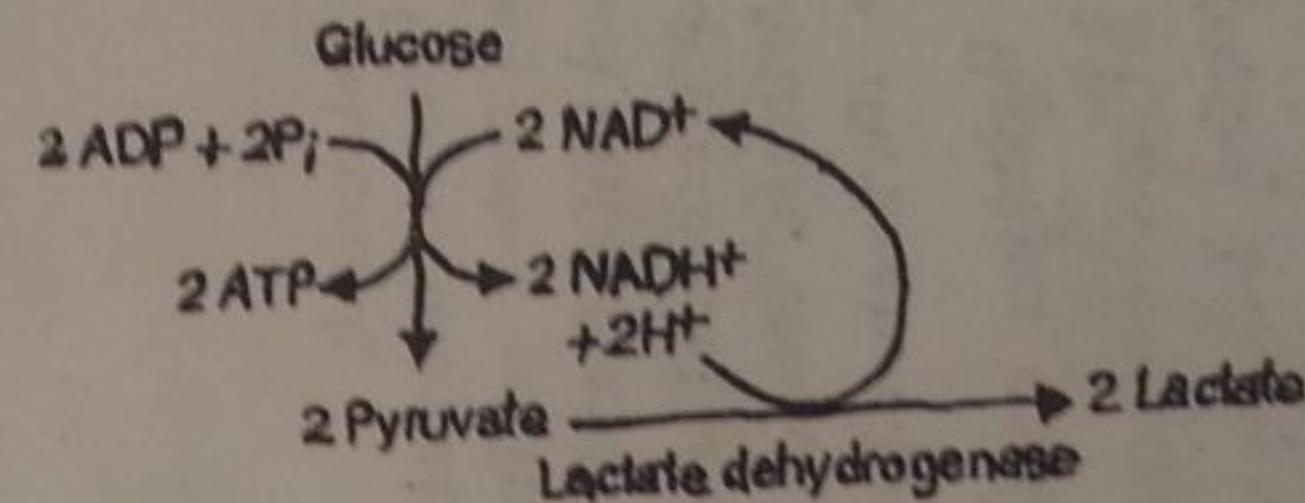
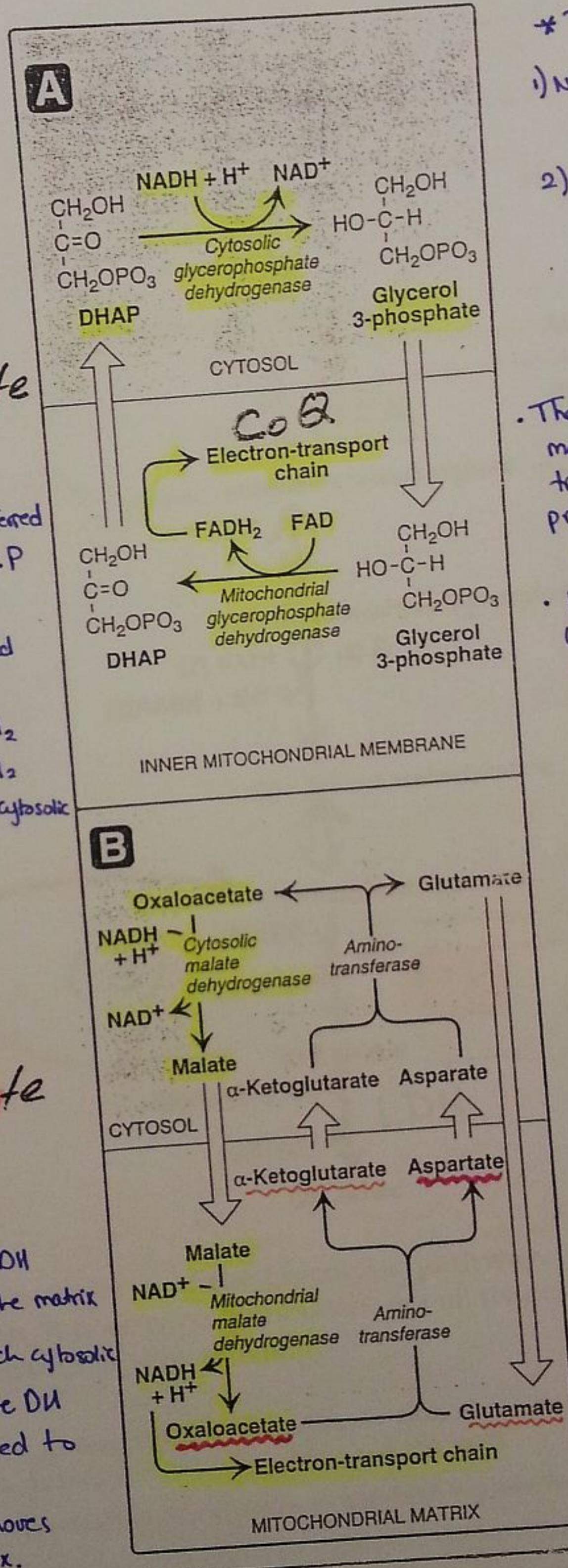


Fig. 22.6. Alternate fates of pyruvate. A. The pyruvate produced by glycolysis enters mitochondria and is oxidized to CO₂ and H₂O. The reducing equivalents in NADH enter mitochondria via a shuttle system. B. Pyruvate is reduced to lactate in the cytosol, thereby using the reducing equivalents in NADH.

11

Shuttle Pathways for the Transport of Electrons across the Inner Mitochondrial Membrane :-



* Glycero-phosphate shuttle

- two electrons are transferred from NADH to dihydroxyacetone P by cytosolic GPDH.
- Glycerol 3P produced is oxidized by mitochondrial isozyme as FAD is reduced to FADH₂.
- CoQ of ETC oxidizes FADH₂.
- two ATPs result from each cytosolic NADH oxidized.

* Malate-Aspartate shuttle

- This shuttle produces NADH instead of FADH₂ in the matrix
- ... it yields 3ATPs for each cytosolic NADH oxidized by malate DH as oxaloacetate is reduced to malate
- a transport protein moves malate into matrix.

* Two major mechanisms
1) NADH-linked conversion of pyruvate → lactate

2) Oxidation of NADH via ETC
→ This requires the malate-aspartate and glycerol 3 phosphate substrate shuttles.

The inner mitochondrial membrane lacks an NADH transporter ∴ NADH produced in cytosol cannot directly enter the matrix.

• However, two electrons (reducing equivalents) of NADH are transported using shuttles.